# Ph.D. COURSE SYLLABUS 2013-2014DEPARTMENT OF ZOOLOGY



School of Biological Sciences

Dr. H. S. Gour University Sagar

Sagar 470003 M.P.

# Ph.D. Course

## **Minimum Total Credits: 80**

Credits through course work each Semester Minimum = 24

➤ 4 Courses X 4 Credits = 16 Credits through course work

+ 08 Credits from research wok each semester

Total Course work Credits = 48 Credits in two semester

Credits through Research (minimum) = 32 after two Semesters

➤ Minimum number of Courses = 06

➤ Minimum Residence = 5 Semester

➤ Maximum Duration = 6 years

## **DEPARTMENT OF ZOOLOGY**

## SCHOOL OF BIOLOGICAL SCIENCES

# DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### I Semester

		Credits
Course I	Code – Zoo C 141: Methodology	04
Course II	Code – Zoo C 142: Modern Instrumentation	04
Course III	Code – Zoo C 143: Research Techniques	04
Course IV	Code – Zoo C 144: Tissue Culture	04
	Research wok	08

#### **II Semester**

Recent topics related to modern researches:

		Credits
Course I	Code – Zoo C 241: Cell Biology	04
Course II	Code – Zoo C 242: Entomology	04
Course III	Code – Zoo C 243: Environmental Biology	04
Course IV	Code – Zoo C 244: Fish & Fisheries	04
	Research wok	08

**Total credits: 48 credits in two semesters** 

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## DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### **I Semester**

Course I Code – Zoo C 141 Course Title: Methodology

L T P C

4 0 0 4

Instruc	tor In-Cl	harge: P	rof. M. Bhide	Lectures
1.	Procure	ement &	Rearing of experimental animal according to animal ethics.	01
2.	Toxicar	nt & Toxi	city.	01
3.	Anaton	ny of sel	ected invertebrates and vertebrates: Alimentary Canal, Excretory	& Nervous
	system	, Brain, N	Nerves – Spinal & cranial, Reproductive system – male & female.	01
4.	Localiza	ation and	d structure of endocrine glands.	01
5.	Experin	nents to	decide sub lethal, lethal & lethal doses, KD0, KD50, KD100, ED0, ED5	o, ED <sub>100</sub> , LD <sub>0</sub> ,
	LD <sub>50</sub> , L[	D <sub>100</sub> , LC <sub>0</sub> ,	LC <sub>50</sub> , LC <sub>100</sub> doses.	01
6.	Osmiur	m ethyl g	gallate method for cytology.	01
7.	Microto	omy		01
	a.	Fixative	es & preservatives	
	b.	Prepara	ation of paraffin blocks of tissue for microtomy	
	c.	Section	cutting & spreading etc.	
8.	Histolo	gical stu	dies: Double staining (Haematoxylin & Eosin).	01
9.	Histoch	nemical s	studies.	
	a.	Prepara	ation of different types of stains and different staining methods:	01
		i.	Mercuric bromophenol blue: for detection of proteins in the tiss	sue.
		ii.	Sudan Black 'B': for detection of lipid in tissue eg. Fat bodies & o	ovaries.
		iii.	Methyl green & Pyronin: to detect DNA & RNA in tissues.	
	b.	Prepara	ation for vital staining:	01

i. Janus green staining of Buccal epithelium for demonstration of mitochondria.

- **ii.** Aceto carmine / aceto orcein staining for polytene chromosomes in salivary gland of Dipteran insects.
- iii. Demonstration of mitosis in squash preparation

**10.** Observations & Results.

01

11. MicroPhotography & preparation of prints.

01

Total Lectures =12

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## DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### I Semester

Course II Code – Zoo C 142 Course Title: Research Techniques

L	Т	Р	С
4	0	0	4

Instructor In-Charge: Prof. S. Banerjee		Lectures
1.	pH determination	01
2.	Spectrometry	01
3.	Chromatography	01
4.	Electrophoresis	01
5.	NMR	01
6.	Centrifugation	01
7.	Phase contrast optics	01
8.	Microscopic techniques	01
9.	Blotting Techniques	01
10.	ELISA	01
11.	Flow cytometery	02

Total Lectures = 12

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## DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### I Semester

Course III Code – Zoo C 143 Course Title: Techniques in Molecular Biology

L	Т	Ρ	U
4	0	0	4

Instruct	Instructor In-Charge: Dr. S.K. Jain and Dr. V. Sharma		
1.	Extraction, purification and separation of: Proteins, DNA and RNA.	01	
2.	Quantitative analysis of Proteins, Sterols, DNA, Vitamins and Carbohydrate.	01	
3.	Literature survey preparation of review articles and research paper.	03	
4.	Antisense and ribozyme technology.	01	
5.	Strategies for designing ribozymes.	01	
6.	Applications of antisense & ribozyme technology.	01	
7.	Molecular mapping of genome.	01	
8.	In situ hybridization for genome analysis.	01	
9.	PCR: Amplification of gene.	01	
10.	BLAST & FASTA.	01	

Total Lectures = 12

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# DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### I Semester

Course IV Code – Zoo C 144 Course Title: Tissue Culture

L T P C 4 0 0 4

Instructor In-Charge: Dr. M. Sikdar		
1.	Bio – safety measures.	01
2.		01
3.	Sterilization methods.	02
4.	Bacterial culture.	01
5.	. Fungal culture.	
6.	Plant cell and tissue culture.	02
	a. Primary culture.	
	b. Secondary culture.	
7.	Animal cell and tissue culture.	02
	a. Primary culture.	
	b. Secondary culture.	
8.	Cryopreservation of cells.	02

**Total Lectures = 12** 

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#### DR. H. S. GOUR UNIVERSITY SAGAR, 470003 (M.P.)

#### II Semester

Course I Code – Zoo C 241 Course Title: Cell Biology

L	Т	Р	С
4	0	0	4

Instructor In-Charge: Dr. S.K.Jain Lectures

- Structure of prokaryotic and eukaryotic cells isolation and growth of cells, cell wall and cell membrane synthesis.
- Microscopic techniques for study of cells and Sub cellular fractionation and criteria of functional integrity.
- 3. Transport of nutrients, ions and macromolecules across membranes. 01
- Cellular response to environmental signals in plants and animals, mechanism of signal transduction.
- 5. Metabolite pathways and their regulation. 01
- Cellular basis of differentiation and development, spatial and temporal regulation of gene expression.
- 7. Protein localization: Synthesis of secretory and membrane proteins, import into nucleus, mitochondria, chloroplast and peroxisomes, receptor mediated endocytosis. 01
- Molecular mechanism of antisense molecules, inhibition of splicing, polyadenylation and translation, disruption of RNA structure and capping, biochemistry of ribozyme,
   Hammer-head, hairpin and other ribozyme.
- 9. Strategies for designing ribozymes and applications of antisense and ribozyme technologies.

- Molecular mapping and map-based cloning, choice of mapping population, simple sequence repeat loci, Southern and fluorescence in situ hybridization for genome analysis, chromosome microdissection and microcloning, molecular markers in genome analysis: RFLP, RAPD and AFLP analysis.
- 11. Molecular markers linked to disease resistance genes, application of RFLP in forensic, disease prognosis, genetic counseling, pedigree varietal etc. Animal trafficking and poaching: Germplasm maintenance, taxonomy and biodiversity.02

**Total Lectures = 12** 

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#### **II Semester**

Course II Code – Zoo C 242 Course Title: Entomology

L	Т	Р	C
4	0	0	4

Instruc	ctor In-Charge: Prof. J. D. Ahi	Lectures
1.	Insect taxonomy & Biodiversity.	01
2.	Insect physiology (Especially reproductive physiology).	01
3.	Insect biochemistry.	01
4.	Insect neuroendocrine organs and insect hormones.	01
5.	Insect immunology.	01
6.	Insect haemolymph proteins.	01
7.	Insect pests.	02
	a. Vegetable pests.	
	b. Stored grain pests.	
	c. Fruit pests.	
8.	Insect pests managements & IPM	01
9.	Insect markers and forensic entomology.	01
10	). Forest entomology.	01
11	Insect hormones.	01

**Total Lectures = 12** 

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#### **II Semester**

Course III Code – Zoo C 243 Course Title: Environmental Biology

L	Т	Р	С
4	0	0	4

Instructor In-Charge: Prof. U. S. Gupta			
1.	Faunal diversity & population density.	01	
2.	Freshwater ecosystem:	01	
	a. Abiotic components.		
	b. Biotic components.		
3.	Food chain, food webs, interaction, energy flow & pyramid.	01	
4.	Productivity.	01	
5.	Toxicants & toxicity.	01	
6.	Vermitechnology & vermicomposting.	01	
7.	Water analysis.	01	
8.	Correlation of abiotic components with faunal diversity & population density.	01	
9.	Adaptation.	01	
10.	Mathematical models in ecology.	01	
11.	Remote sensing.	01	
12.	Waste water treatment.	01	

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#### **II Semester**

Course IV Code – Zoo C 244 Course Title: Fish & Fisheries

L	Т	Р	С
4	0	0	4

	Instruc	tor In-Charge: Prof. D. K. Saraf	Lectures		
1.	Outline	e of classification of fish.	01		
2.	Fish ide	entification, purpose and importance of aquaculture.	01		
3.	Basic c	onsideration in the selection of species for culture, cultivable fresh water fish	es. 01		
4.	Cultivable fresh water fin fishes (food and feeding habits and biology of carps, air breathing and				
	cat fish	nes)	01		
5.	Food, f	eeding habits and factors influencing feeding, reproduction.	01		
6.	Artificial propagation.				
	a.	Structure of ovary, maturity, stages of ovary.			
	b.	Structure of testes, maturity, stages of testes.			
	c.	Fecundity.			
	d.	Relation between endocrine and reproduction.			
7.	Induced breeding.		01		
8.	Genetic improvement of fish stock. 01				
9.	. Hybridization, sex manipulation, chromosomal manipulation.				
10.	10. Gene engineering, genetic marker, genetic conservation in fish. 01				
11.	11. Integrated fish culture. 01				

**Total Lectures = 12**